

Diagnosis of Shashi-Pena Syndrome Caused by Chromosomal Rearrangement Using Nanopore Sequencing

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Abstract

Background and Objectives

The aim of this study was to uncover the genetic cause of delayed psychomotor development and variable intellectual disability in a proband whose previous genetic analyses, including chromosome microarray and whole exome sequencing, had been negative.

Methods

Long-read sequencing Oxford Nanopore Technology and RNA-seq analysis were performed on peripheral blood mononuclear cells. Genes with a fold change ≥ 1.5 and $p \leq 0.05$ were identified as differentially expressed.

Results

Clinical examinations showed that the proband's features were similar to a rare autosomal-dominant neurodevelopmental syndrome, Shashi-Pena syndrome (MIM #617190). Karyotyping showed that a chromosomal balanced translocation t(2; 11) (p23; q23) was detected in the proband, her father, and her grandmother. Meanwhile, long-read sequencing identified 102 balanced translocations and 145 inversions affecting *ASXL2* at an average of 15 \times . Combined with the family's RNA-seq results, the average mRNA expression of *ASXL2* decreased in the patients.

Discussion

We identified a complex chromosomal rearrangement affecting *ASXL2* as a pathogenic mechanism of Shashi-Pena syndrome in a Chinese family. This case study suggests that nanopore sequencing is suitable for pathogenic analysis of complex rearrangements, providing new avenues for the diagnosis of genetic diseases.

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Glossary

CMA = Chromosome Microarray; **ONT** = Oxford Nanopore Technology; **SMRT** = PacBio Single-molecule Real-time Sequencing; **ID** = Intellectual Disability; **SVs** = Structural variation; **TRA** = Balanced Translocations.

The *ASXL* (additional sex combs-like) gene family of epigenetic regulators including *ASXL1* [MIM: 612990], *ASXL2* [MIM: 612991], and *ASXL3* [MIM: 615115] code for polycomb proteins that act as histone methyltransferases,¹ serve as epigenetic scaffolding proteins,² and participate in body patterning.³ *ASXL1* gene mutations are frequently detected in myeloid neoplasms^{4,5} and cause Bohring–Opitz syndrome.⁶ *ASXL3* gene mutations result in Bainbridge–Ropers syndrome (MIM #615485).⁷ *ASXL2* is conserved in *Drosophila*, mice, and humans and is part of the transcription regulation complex.⁸ However, *ASXL2* mutations are rarely reported in humans.

Previous reports have shown that *ASXL2* mutants in 6 patients caused Shashi-Pena syndrome.³ All 6 had de novo truncating variants in *ASXL2*. The common clinical features in Shashi-Pena syndrome are neurologic and neurodevelopmental disorders with some facial features, whereas differences in the clinical phenotypes for each individual could be found.

Long-read sequencing technologies, such as Oxford Nanopore Technologies (ONTs)⁹ and PacBio single-molecule real-time sequencing,¹⁰ offer complementary strengths to traditional whole exome/whole genome sequencing based on short-read sequencing. Long-read sequencing can produce read lengths (typically 1–150 kb or longer) that are far higher than the ~150 bp produced by short-read sequencing, therefore allowing for the resolution of breakpoints of complex structural variations (SVs),¹¹ the detection of long tandem repeats,¹² or DNA methylation on N⁶-adenine.¹³ Balanced translocations (TRA), a type of SVs, involve the exchange of genomic regions between nonhomologous chromosomes without the gain or loss of genetic material. Most TRA carriers are phenotypically normal, but the pathogenesis of abnormal TRA carriers remains unclear because of the technical limitations of detection. However, an association with abnormal phenotypes, including intellectual disability, developmental delay, and other congenital abnormalities, has been estimated in 6%–9% of de novo cases.¹⁴

We used ONTs to perform a genetic study of a proband (III-1) with developmental retardation, for whom a balanced translocation t(2; 11) (p23; q23) was identified by a chromosome karyotypic analysis, and negative results were found by chromosomal microarray analysis and whole exome sequencing. Moreover, RNA sequencing was carried out on peripheral blood of the whole family. By combining the results of the breakpoint candidate genes, RNA-seq, and qPCR, we concluded that a complex rearrangement of the chromosomes decreased the mRNA expression of *ASXL2* in the patients and led to a rare autosomal-dominant neurodevelopmental syndrome, Shashi-Pena syndrome.

Methods

Standard Protocol Approvals, Registrations, and Patient Consents

The samples of peripheral blood were taken from each patient on the same day. All procedures performed in studies involving human participants were in accordance with the Declaration of Helsinki and its later amendments or comparable ethical standards, with written informed consent obtained from all patients and their parents/guardians. The Research Ethics Committee of Nanjing Maternal and Child Health Hospital approved the study ([2019] KY-081).

Study Design and Data Collection

The proband is a 6-year-old girl who was admitted to our hospital at 3 years old because of speech delay. Her karyotype is t(2; 11) (p23; q23). Data were collected on her family history, medical history, and medications. The proband underwent a physical examination including length, weight, occipitofrontal head circumference, additional biochemical assays, Wechsler Intelligence test, Peabody test, X-ray examinations, and brain MRI.

Molecular Testing and Variant Curation

A DNA sample of the proband was sequenced using the Nanopore PromethION sequencer (ONTs, UK). Genomic DNA was extracted by QIAGEN genomic DNA extraction kit (Cat#13323, QIAGEN). The extracted DNA was detected by NanoDrop 1 UV-Vis spectrophotometer (Thermo Fisher Scientific) for DNA purity (OD 260/280 ranging from 1.8 to 2.0 and OD 260/230 between 2.0 and 2.2); then a Qubit 3.0 Fluorometer (Invitrogen) was used to quantify DNA accurately. Long DNA fragments were size selected using the BluePippin system (Sage Science), and then sequencing adapters supplied in the SQK-LSK109 kit were attached to the DNA ends. Raw data collected as FAST5 files were converted to FASTQ format using guppy 4.5.3 (ONTs). Base-called data passing quality control (quality score ≥ 9) was aligned to the hg19 human reference genome using Minimap2 (version 2.17) (github.com/lh3/minimap2). SVs were called with Sniffles (version 1.0.11) (github.com/fritzsedlazeck/Sniffles), and the major types of SVs included deletion, insertion, duplication, inversion, and TRA. Further annotation of SVs was carried out using ANNOVAR¹⁵ (github.com/WGLab/doc-ANNOVAR), combining public databases such as 1,000 Genome Phase 3,¹⁶ DGV gold standard CNV,^{17,18} dbVar nstd37,¹⁸ and DECIPHER.¹⁹

PCR primers were designed to detect the translocation breakpoints for this family. Primer3web (version 4.1.0, primer3.ut.ee/) was used for primer design. The sequences of

Figure 1 Clinical Signs and Brain MRI of the Patient With an *ASXL2* Mutation



(A) The pedigree of three family members with Shashi-Pena Syndrome and the marriage status of the proband's parents (II - 1 and II - 2). Black arrow, proband; filled symbol, patient with Shashi-Pena Syndrome. (B) Proband at 5 years 11 months. Note the hypertelorism, arched eyebrows, and abnormal shape of the ears. (C) Brain MRI of the patient at 5 years 11 months of age. Axial image shows periventricular leukomalacia. Sagittal image shows myelination delay (white arrow) and thin corpus callosum (white arrowhead).

the breakpoint PCR primers in this study are listed in eTable 1, links.lww.com/NXG/A487. PCR products were electrophoresed on a 1.0% agarose gel, and the single, clear bands from normal and derivative chromosomes were further analyzed by Sanger sequencing on an ABI3730XL sequencer (Thermo Fisher Scientific).

Total RNA was extracted from the peripheral blood of the family members (I-1, I-2, II-2, and III-1), according to the manufacturer's specifications. The primer sequences were designed in the laboratory and synthesized by TsingKe Biotech based on the mRNA sequences obtained from the NCBI database in eTable 1, links.lww.com/NXG/A487. The expression levels of the mRNAs were normalized to ACTB and were calculated using the $2^{-\Delta\Delta C_t}$ method, a convenient way to analyze the relative changes in gene expression from real-time quantitative PCR experiments.

Statistical Analysis

We used the *t* test to detect the mRNA expression difference between patients and control. Values of $p < 0.05$ were considered statistically significant.

Data Availability

The data sets used and analyzed during this study are available from the corresponding authors upon reasonable request.

Results

Patient Characteristics and Clinical Tests

The proband (III-1) is a 6-year-old girl who came to our attention at the age of 3 because of speech delay (F). She was born at 39⁺⁵ weeks of gestation without any complications via Cesarean operation. Her birth length was 49 cm, and her birth weight was 2,715 g. The Apgar score was 6 points (color: acrocyanotic; heart rate: >100/min; reflex irritability: grimace; muscle tone: some flexion; respiration: weak cry, hypoventilation) in the first minute and 8 points (color: completely

pink; heart rate: >100/min; reflex irritability: grimace; muscle tone: some flexion; respiration: good, crying) in the first 5 minutes. She was sent to the NICU because of neonatal feeding difficulties (HP_0011968) and pneumonia in the first week.

The recognizable pattern of features and the corresponding Human Phenotype Ontology terms seen in the proband (III-1) include short stature (HP_0008848), arched eyebrows (HP_0002553), hypertelorism (HP_0000316), and low-set abnormally shaped ears (HP_0000369) (Figure 1B, Table 1). Examination of her growth parameters showed severe growth retardation during the first 5 years 11 months (weight < third percentile, length < 15th percentile, height < 15th percentile, occipitofrontal head circumference < third percentile). Her developmental delay (HP_0001263) was severe, and the intellectual disability (HP_0001249) and speech delay were moderate via the Wechsler Intelligence test and Peabody test (GMQ 76/P5, FMQ 85/P16, TMQ 78/P7, and SM:8, IQ = 62). A wrist X-ray examination showed osteoporosis. Her heart showed mild mitral regurgitation and mild tricuspid regurgitation (HP_0030680) on an echocardiograph examination. Left cerebral ventriculomegaly, myelination delay, thin corpus callosum, and periventricular leukomalacia were found, and the width of the posterior angle of the left lateral ventricle was ~1.17 cm on brain MRI (Figure 1C). Additional biochemical assays were performed on the proband. Her fasting blood glucose value, levels of cholesterol, triglycerides, and chlorine, thyroid-stimulating hormone, free triiodothyronine (FT3), free thyroxine (FT4), growth hormone, insulin, serum iron, serum copper, ceruloplasmin, serum zinc, serum magnesium, and serum calcium were at normal levels. An investigation of liver function showed that the aminotransferases (aspartate aminotransferase and alanine aminotransferase) and other biochemistry were normal.

A further study was conducted on the family members to trace the source of the complex TRA. Peripheral blood DNA of the family (I-1, I-2, and II-2) was collected, and a balanced translocation t(2; 11) (p23; q23) was identified by chromosome karyotyping in I-1 and II-2. The results of PCR and

Table 1 Clinical Features From Affected Members in the Family Comparison to *ASXL2*-Associated Disorders in Previous Report

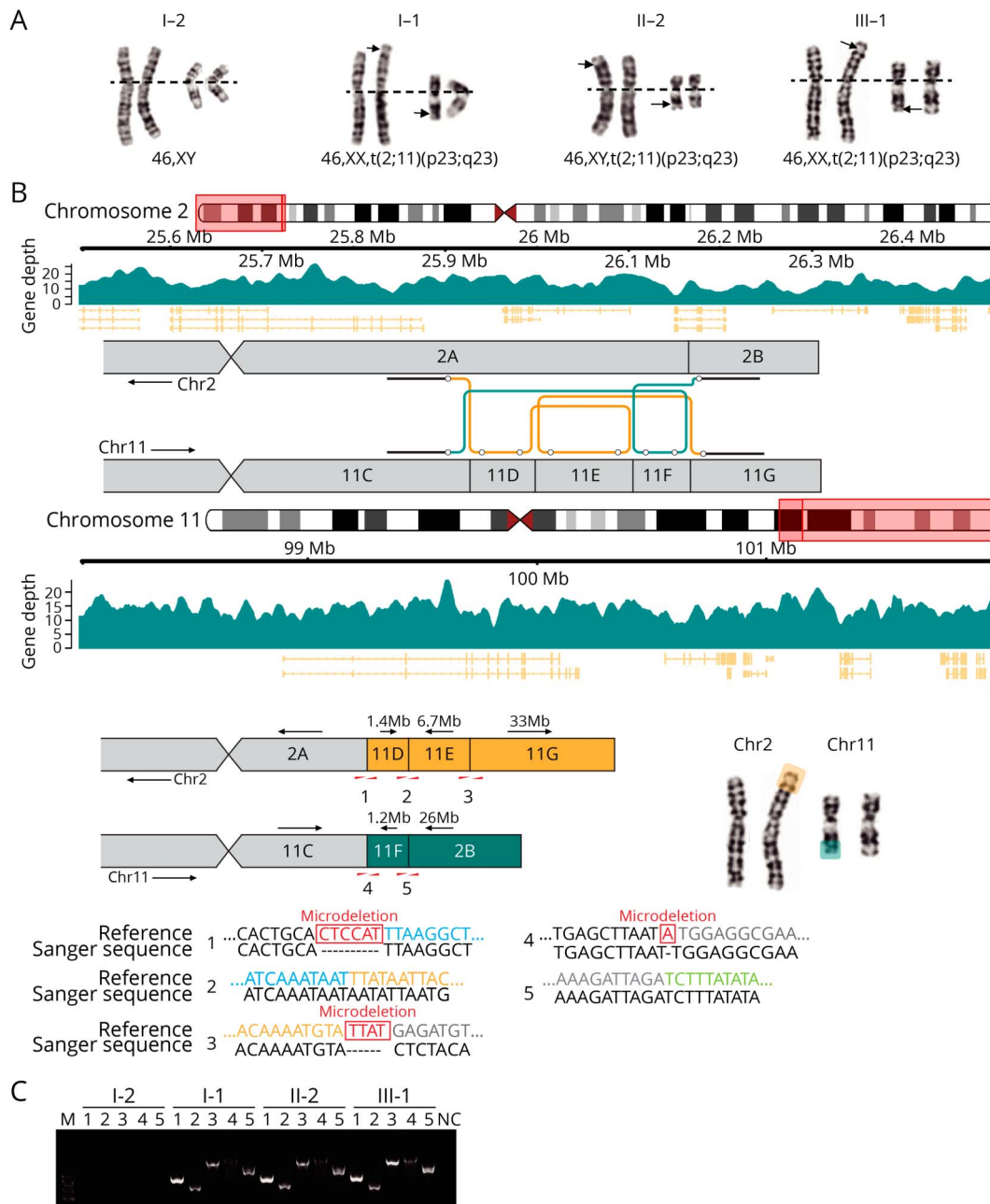
Clinical findings	III-1	II-2	I-1	<i>ASXL2</i> mutation in previous report ³
Facial features	Hypertelorism, arched eyebrows	Arched eyebrows	Normal	Hypertelorism, arched eyebrows, prominent eyes, ptosis of eyelids, epicanthal folds, prominent glabella, synophrys, small upper vermilion, and broad nasal tip
Ears	Low-set	Normal	Normal	Posteriorly rotated, low-set, cupped, overfolded, and thick ear lobes.
Growth parameters at last examination	At 5 y 11 mo: severe growth retardation			
Weight <3rd percentile,				
Length/height <15th percentile,				
Occipitofrontal head circumference <3rd percentile	Normal	Normal	Normal height and weight, occipitofrontal head circumference >97th percentile (macrocephaly)	
Feeding difficulties	Present only shortly after birth	Present only shortly after birth	No record	Present only in neonatal period
Hypotonia	None	None	None	Present
Episodes of hypoglycemia	None	None	None	Has required continuous feeding since neonatal period, episodic, starting at 2.5 y of age
Seizures	None	None	None	Febrile and nonfebrile, suspected
Skin	Normal	Normal	Normal	Glabellar nevus flammeus; capillary malformations on trunk, neck, and behind ears; deep palmar creases
Developmental delay	Severe	Normal	Normal	Severe
Intellectual disability	Moderate	Mild	Normal	Severe
Delayed speech	Moderate	Normal	Normal	Severe
Peabody	GMQ 76/P5, FMQ 85/P16, TMQ 78/P7	Not available	Not available	Not available
Wechsler intelligence test	SM: 8, IQ = 62	Not available	Not available	Not available
Congenital heart disease	Mild mitral regurgitation, mild tricuspid regurgitation	Normal	Normal	Atrial septal defect, patent ductus arteriosus, left ventricular dysfunction
Brain MRI	Myelination delay, thin corpus callosum and PVL, left cerebral ventriculomegaly ~1.17 cm	Not available	Not available	Increased extra-axial cerebral space, white-matter volume loss, increased extra-axial cerebral space, choroid plexus papilloma,
Skeletal and/or extremity manifestations	Osteopenia	Normal	Normal	Increased density of alveolar bone, advanced bone age, kyphosis, scoliosis, multiple bilateral fractures, overlappings, and fourth toes

Abbreviation: PVL = periventricular leukomalacia.

Sanger sequencing showed that these sequences were the same in I-1, II-2, and III-1. The karyotype of the proband is t(2; 11) (p23; q23), and this TRA was inherited from her grandmother (Figure 2A).

The father (II-2) was 172 cm tall and 74 kg at the age of 35, and the clinical features include arched eyebrows (HP_0002553), feeding difficulties (HP_0011968), and moderate verbal comprehension during communication. No bruising, cuts, or lumps were found in

Figure 2 Karyotype Analysis and Composition of the Derivative Chromosomes in a Balanced Translocation, With the Precise Coordinates of Breakpoints Detected and Confirmed by Sanger Sequencing



(A) Conventional G-banded partial karyotypes in the family members. (B) The “Subway” plot Shows how each region is connected and allows for reconstruction of the new DNA sequence and gene order in the individual, and the subsequent karyotype reveals a complex translocation involving chromosomes 2 and 11. Breakpoints were confirmed by Sanger sequencing, and microdeletions were detected. Primers 1, 2, 3, 4, and 5 (red arrow heads). (C) PCR validation shows that the variant was caused by balanced translocation in the family (I-1, I-2, II-2, and III-1). No bands were detected in the proband’s grandfather (I-2). PCR products of three patients and one normal control are shown (DL2,000 DNA Marker).

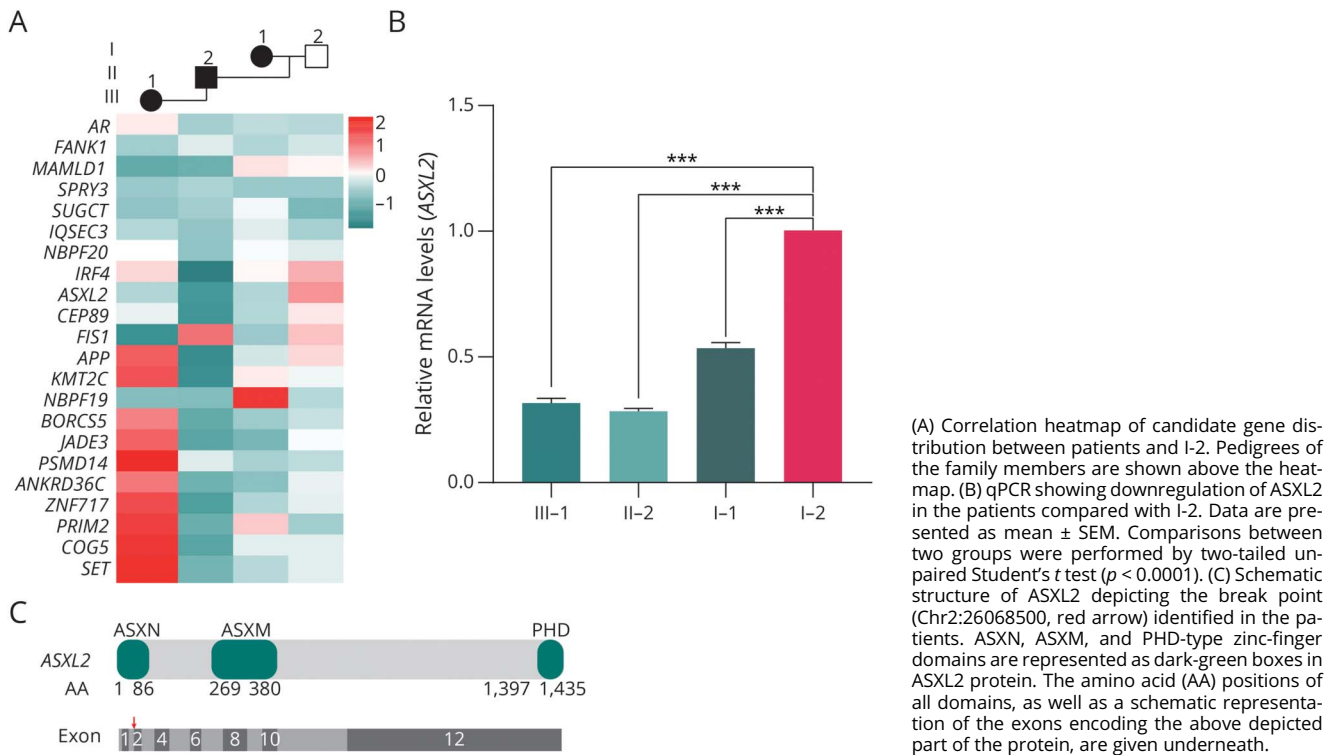
his skin; seizures and episodes of hypoglycemia have never appeared, and he was movement barrier-free as found by visual examination. The grandmother was 156 cm tall and 57.3 kg at 65 years old, and she was thin and short in childhood. No redness, swelling, deformity, and movement barrier were found by visual examination, and seizures and episodes of hypoglycemia have never appeared. Developmental delay and intellectual disability are not known in her childhood, but her talk and verbal comprehension are

normal. In summary, the phenotypes of the 3 patients are similar to Shashi-Pena syndrome caused by *ASXL2* mutants (Table 1).

Complex Chromosome Rearrangements Found by Nanopore Sequencing

Next, we performed whole genome long-read sequencing analysis on the proband (III-1) and used different analytical strategies and tools to analyze the translocation breakpoints.

Figure 3 Expression of Candidate Genes in Patients and the Control



SVs including 102 translocations and 145 inversions were found in the proband (III-1) at an average of 15× of the whole genome, and the balanced translocation was confirmed as t(2; 11) (p23.3; q22.1) based on the karyotyping results. Our analysis workflow successfully detected the complex SV breakpoints in the proband (III-1) and confirmed the karyotype as 46,XX,t(2; 11) (p23; q23)pat.Seq[GRCh37]der(2)t(2; 11) (p23.3; q22.1)del(q22.1)inv(11) (q22.1),der(11)t(2; 11) (p23.3; q22.1),ins(11) (q22.1)[chr11:pter_101817177:chr11:99900659_100571704:chr11:99900660_98496146:chr2:26068500_cen_qter].[chr11:pter_cen_98496149:chr11:101817176_100571719:chr2:26068500_qter] (Figure 2B). The coordinates of the predicted breakpoints that correlated with the cytogenetic report and the nanopore results showed the extent of concordance of the predicted breakpoints with the Sanger sequencing results (Figure 2C). There were discrepancies of a few bases because of microdeletions in all the samples, and Sanger sequencing confirmed the microdeletions.

Complex chromosome rearrangements affecting ASXL2 as a pathogenic mechanism of Shashi-Pena syndrome.

In our target regions, an ONT analysis showed 3 breakpoints in the complex balanced translocation and inversions affecting ASXL2 (eTable 2, links.lww.com/NXG/A487). The RNA-seq was conducted on peripheral blood of the family members, and the average mRNA expression of ASXL2 decreased among the patients (*t* test, $p = 0.0387$) (Figure 3A). Furthermore, the

qPCR showed that the mRNA expression of ASXL2 decreased in the patients, consistent with the RNA-seq results (unpaired *t* test, $p < 0.0001$) (Figure 3B). In conclusion, truncating variants in ASXL2 via a complex TRA t(2; 11) (p23.3; q22.1) underlie Shashi-Pena syndrome (MIM:617,190) with a clinically recognizable phenotype (Figure 3C).

In summary, the karyotype t(2; 11) (p23.3; q22.1) divides ASXL2 into 2 parts, affecting its expression and leading to Shashi-Pena syndrome in a Chinese family. This case study suggests that nanopore sequencing is suitable for the pathogenic analysis of complex rearrangements, providing new avenues for the diagnosis of agnogenic diseases.

Discussion

Genomic SVs, including translocations, inversions, insertions, deletions, and duplications, are challenging to detect reliably using traditional genomic technologies.²⁰⁻²² In particular, balanced translocations and inversions cannot be identified by microarrays or by short-read sequencing.²³ The precise localization of breakpoints is vital for exploring genetic causes in patients with balanced translocations or inversions.²⁴ Long-read sequencing techniques may detect these structural variants in a more direct, efficient, and accurate manner.²⁵

In this study, we identified 3 patients in a family with a complex SV, t(2; 11) with breakpoints at p23.3 and q22.1, and

identified possible candidate genes by Oxford Nanopore Technology. Using RNA-seq and qPCR, we found that the breakpoints disrupted *ASXL2* gene expression and finally concluded that a complex rearrangement of the chromosomes decreased the mRNA expression of *ASXL2* in the patients and led to a rare autosomal-dominant neurodevelopmental syndrome, Shashi-Pena syndrome. Shashi-Pena syndrome, reported in 2016 in the United States,³ is caused by *ASXL2* mutations and shows the common clinical features of developmental disabilities, intellectual disability, delayed speech, and some facial features. However, differences were found between cases, such as the level of intellectual disability, congenital heart disease, and brain MRI, which may be because of different mutation types or incomplete penetrance.

We report a case of Shashi-Pena syndrome caused by a balanced translocation in a Chinese family, which expands the mutation spectrum of *ASXL2* and increases the number of reported patients, and the results could allow better counseling for patients. More clinical reports of individuals with damaging *ASXL2* variants and related clinical features will provide further clarity in the future.

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Disclosure

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Jianxin Tan, PhD	Nanjing Maternity and Child Health Care Hospital, Nanjing, China	Clinical genetic evaluation and reviewed the manuscript

Appendix (continued)

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Cuiping Zhang, MD	Nanjing Maternity and Child Health Care Hospital, Nanjing, China	Molecular data acquisition
Jing Zhou, MD	Nanjing Maternity and Child Health Care Hospital, Nanjing, China	Molecular data acquisition
Ping Hu, PhD	Nanjing Maternity and Child Health Care Hospital, Nanjing, China	Funded the project and supervised and managed the study
Zhengfeng Xu, MD	Nanjing Maternity and Child Health Care Hospital, Nanjing, China	Funded the project, interpreted the data, and drafted the manuscript

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