Possible Somatic Mosaicism of Novel *FUS* Variant in Familial Amyotrophic Lateral Sclerosis

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Fused-in-sarcoma (*FUS*) mutation is associated with a familial amyotrophic lateral sclerosis (fALS) characterized by early onset and rapid progression. Here, the authors describe a rare case of possible somatic mosaicism of a novel *FUS* variant for a fALS family with intrafamilial clinical variability.

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Page e555

Cases

Case 1

The index case (II-6, figure, A) was a woman who had gradual weight loss and gait disorder from age 44 years. Two years later, she could not walk without assistance. Neurologic examination at age 46 revealed tongue atrophy and fasciculation, together with weakness and fasciculation in her limbs and trunk. Pathologic reflexes were observed bilaterally. Needle EMG showed active and chronic neurologic changes in multiple regions, suggesting ALS. Enteral nutrition with percutaneous endoscopic gastrostomy were started at age 47, and artificial ventilation with tracheostomy at age 50.

Case 2

The son of the index case (III-5, figure, A) was born by full-term normal delivery. His psychomotor milestones were normal. At age 11 years, he had difficulty standing from a squatting position. At age 12, he was seen by a pediatrician because of his inability to walk. Neurologic examination indicated muscle weakness in his limbs. Increased deep tendon reflexes were observed. Muscle biopsy was performed twice, with results indicating neurogenic diseases including ALS rather than myopathies. He complained of respiratory distress 2 months later and required tracheostomy and artificial ventilation at 6 months after admission. Six months later, he was in a complete locked-in state. He died at the age of 33.

Whole exome analysis using a blood sample from Case 2 identified a novel heterozygous *FUS* variant (c. 1,542_1545delGGGT, p.Gly515Serfs13*) (figure, A and B). Surprisingly, it was difficult to detect the same variant in the blood sample of case 1 because of the low frequency of the mutant allele; only 1/30 reads counted using the Integrative Genomics Viewer (figure e-1, links.lww.com/NXG/A363). The targeted next generation sequencing (NGS) data from her peripheral blood and saliva showed repeatable low variant rates of 4/327 and 5/280 reads, respectively. Sanger sequencing confirmed that the mother had lower mutant allele fraction in blood than her son (figure, B). Furthermore, higher mutant allele fractions were detected in her hair and nail than blood and saliva (figure, B).

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Figure Somatic Mosaicism of *FUS* Variant Causing Familial Amyotrophic Lateral Sclerosis With Different Clinical Courses Within the Family (A) Pedigree of the Family

Black symbols indicate individuals affected by amyotrophic lateral sclerosis. Asterisks indicate individuals whose DNA was studied. Case 1 (II-6) is the index patient (arrow) with mosaic variant of *FUS*. (B) Sanger sequencing of DNA from peripheral blood of the unaffected father (II-5) shows wild type. Sanger sequencing of DNA from peripheral blood of case 2 (III-5) clearly shows a heterozygous *FUS* variant (c.1542_1545delGGGT, p.Gly515Serfs13*). Sanger sequencing of DNA from peripheral blood, saliva, hair, and nail samples of case 1 (II-6) confirms different frequencies of *FUS* mutant allele in various tissues. Black arrows indicate start of deletion. Sequencing results are shown in reverse. Supplementary data and table, links.lww.com/NXG/A369 contain additional details on the methodology. *FUS* = *Fused-in-sarcoma*.

Discussion

We present a rare case of a possible somatic and germline mosaic variant in a patient with fALS. Sequencing confirmed lower mutant allele fraction in blood cells of the index case than her son. The index case also had different mutant allele fractions in various tissues. These observations indicate that she has a somatic variant of *FUS* across multiple tissues, which affects germline and somatic cells. This variant may have arisen before the 3 germ layers split, which could lead to mosaicism not restricted to the ectoderm, with lower but equivalent proportion of variant both in the mesoderm-derived blood and mesoderm- and ectoderm-derived saliva. Whether she had mosaicism in the CNS could not be investigated. However, given the higher variant fractions in tissues such as nail from the ectoderm system, a similar tendency may be found in neural tissues. Targeted deep sequencing in trio analysis revealed ~3% of causative de novo mutations display postzygotic mosaicism.¹

Her son showed more severe clinical course with juvenile onset and rapid progression. Recently, we found genetic linkage in 49.5% of a Japanese cohort of familial ALS.² Most of the *FUS*-linked patients with fALS showed phenotype characterized by early onset, in contrast to patients with sporadic ALS.³ The variant p.G515Sfs13* is located in the highly conserved C terminal of *FUS* protein, where most of the *FUS* variants have been identified. The mutant protein has a truncated peptide from 515 to 526 amino acids, which coincides with the mutant terminal peptides affected by p.R495QfsX527 variant.⁴ The degree of *FUS* protein mislocalization from nucleus to cytoplasm by C-terminal deletion correlates with the severity of clinical phenotype.⁵ However, variable phenotypes of the same *FUS* variant have been observed.⁴

Somatic mosaicism in ALS is probably rare, but this possibility should be considered. Several studies indicated somatic variant in other neurodegenerative disorders such as Alzheimer's disease.⁶ The clinical course of case 1 suggests that somatic

variant may have a milder phenotype depending on the extent of mosaicism. In ALS, repeat expansions of *C9ORF72* exhibit both infrafamilial and somatic instability leading to mosaicism.⁷ In patients with intrafamilial variability or apparently sporadic disease, it may be important to suspect somatic mosaic variants during screening for candidate variants using high coverage NGS. Furthermore, variant analysis using multiple tissues other than peripheral blood may help identify low-level mosaicism. Additional studies including histopathology of these cases would contribute to elucidate the pathogenesis of ALS.

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Disclosure

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Shin Hisahara, MD, PhD	Sapporo Medical University, Sapporo, Japan	Designed and conceptualized study and drafted the manuscript
Ayumi Nishiyama, MD, PhD	Tohoku University, Sendai, Japan	Performed analysis, interpreted the data, and drafted the manuscript
Emiko Tsuda, MD, PhD	Sapporo Shirakabadai Hospital, Sapporo, Japan	Provided clinical information

Appendix (continued)

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Syuuichirou Suzuki, MD, PhD	Sapporo Medical University, Sapporo, Japan	Provided clinical information
Akihiro Matsumura, MD, PhD	Sapporo Medical University, Sapporo, Japan	Provided clinical information
Aki Ishikawa, MD, PhD	Sapporo Medical University, Sapporo, Japan	Performed genetic consultation
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Ikuko N. Motoike, PhD	Tohoku Medical Megabank Organization, Sendai, Japan	Performed analysis and acquisition of data
Masashi Aoki, MD, PhD	Tohoku University, Sendai, Japan	Supervised the study and obtained funding
Yoko Aoki, MD, PhD	Tohoku University, Sendai, Japan	Supervised genomic analyses, and critically revised the manuscript
Shun Shimohama, MD, PhD	Sapporo Medical University, Sapporo, Japan	Supervised the study, critically revised the manuscript, and obtained funding

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