Association study between multiple system atrophy and TREM2 p.R47H

Kotaro Ogaki, MD, PhD, Michael G. Heckman, MS, Shunsuke Koga, MD, PhD, Yuka A. Martens, PhD, Catherine Labbé, PhD, Oswaldo Lorenzo-Betancor, MD, PhD, Ronald L. Walton, BSc, Alexandra I. Soto, BSc, Emily R. Vargas, MPH, Shinsuke Fujioka, MD, Ryan J. Uitti, MD, Jay A. van Gerpen, MD, William P. Cheshire, MD, Steven G. Younkin, MD, PhD, Zbigniew K. Wszolek, MD, Phillip A. Low, MD, Wolfgang Singer, MD, Guojun Bu, PhD, Dennis W. Dickson, MD, and Owen A. Ross, PhD

Correspondence
Dr. Ross
ross.owen@mayo.edu

Neurol Genet 2018;4:e257. doi:10.1212/NXG.0000000000000257

Abstract

Objective

The triggering receptor expressed on myeloid cells 2 (TREM2) p.R47H substitution (rs75932628) is a risk factor for Alzheimer disease (AD) but has not been well studied in relation to the risk of multiple system atrophy (MSA); the aim of this study was to evaluate the association between the TREM2 p.R47H variant and the risk of MSA.

Methods

A total of 168 patients with pathologically confirmed MSA, 89 patients with clinically diagnosed MSA, and 1,695 controls were included. TREM2 p.R47H was genotyped and assessed for association with MSA. Positive results in the Taqman genotyping assay were confirmed by Sanger sequencing. The primary comparison involved patients with pathologically confirmed MSA and controls due to the definitive MSA diagnosis in the pathologically confirmed series.

Results

We identified TREM2 p.R47H in 3 patients with pathologically confirmed MSA (1.79%), 1 patient with clinically diagnosed MSA (1.12%), and 7 controls (0.41%). Minimal AD pathology was observed for the pathologically confirmed MSA p.R47H carriers. For the primary comparison of patients with pathologically confirmed MSA and controls, risk of disease was significantly higher for p.R47H carriers (odds ratio [OR]: 4.39, $p = 0.033$). When supplementing the 168 pathologically confirmed patients with the 89 clinically diagnosed and examining the combined MSA series, the association with TREM2 p.R47H remained significant (OR: 3.81, $p = 0.034$).

Conclusions

Our preliminary results suggest that the TREM2 p.R47H substitution may be a risk factor for MSA, implying a link to neuroinflammatory processes, especially microglial activation. Validation of this finding will be important, given our relatively small sample size; meta-analytic approaches will be needed to better define the role of this variant in MSA.

From the Department of Neuroscience (K.O., S.K., Y.A.M., C.L., O.L.-B., R.L.W., A.I.S., S.F., S.G.Y., G.B., D.W.D., O.A.R.), Mayo Clinic, Jacksonville, FL; Department of Neurology (K.O.), Juntendo University Shizuoka Hospital, Izunokunishi, Shizuoka, Japan; Division of Biomedical Statistics and Informatics (M.G.H., E.R.V.), Mayo Clinic, Jacksonville, FL; Department of Neurology (R.J.U., J.A.v.G., W.P.C., Z.K.W.), Mayo Clinic, Jacksonville, FL; Department of Neurology (P.A.L., W.S.), Mayo Clinic, Rochester, MN; Mayo Graduate School (O.A.R.), Neurobiology of Disease, Jacksonville, FL; and Department of Clinical Genomics (O.A.R.), Jacksonville, FL.

Funding information and disclosures are provided at the end of the article. Full disclosure form information provided by the authors is available with the full text of this article at Neurology.org/NG.

The Article Processing Charge was funded by the authors.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND), which permits downloading and sharing the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Glossary

AD = Alzheimer disease; **FTD** = frontotemporal dementia; **MSA** = multiple system atrophy; **MSA-C** = MSA with predominant olivopontocerebellar involvement; **MSA-P** = MSA with predominant striatonigral involvement; **OR** = odds ratio; **PD** = Parkinson disease; **TREM2** = triggering receptor expressed on myeloid cells 2.

Multiple system atrophy (MSA) is a rare, rapidly progressing, adult-onset neurodegenerative disease that is clinically characterized by parkinsonism, autonomic failure, pyramidal symptoms, and cerebellar ataxia and pathologically defined by the presence of α -synuclein-positive glial cytoplasmic inclusions.¹ Uncovering genetic risk factors for MSA has proven difficult because of the challenge of accumulating a reasonable sample size and potential for clinical misdiagnosis. Nonetheless, studies have nominated several genes that may be associated with the risk of MSA.^{1,2} The first genome-wide association study of MSA was recently completed, and although no genome-wide significant associations were identified, suggestive associations were identified involving variants in *MAPT*, *FBXO47*, *ELOVL7*, and *EDN1*.³ Clearly, much remains to be understood regarding the genetics of MSA, and replication of previously nominated MSA susceptibility loci remains crucial.

A relatively rare substitution p.R47H (rs75932628) in the triggering receptor expressed on myeloid cells 2 (TREM2) protein is now a well-established risk factor for Alzheimer disease (AD) and has been suggested to play a role in other neurodegenerative disorders such as frontotemporal dementia (FTD) and Parkinson disease (PD).⁴ Of interest, mutations in the *TREM2* gene were originally reported as a genetic cause of Nasu-Hakola disease, a rare leukodystrophy characterized by progressive presenile dementia and bone cysts.⁴ Given the important role of microglia in neuroinflammation, we evaluated the association between the TREM2 p.R47H variant and the risk of MSA using a predominantly neuropathologically confirmed MSA series.

Methods

Study participants

This study included 168 patients with pathologically confirmed MSA, 89 patients with clinically diagnosed MSA, and 1,695 healthy controls. The pathologically confirmed MSA series consisted of all cases obtained from the Mayo Clinic brain bank for neurodegenerative disorders in Jacksonville, FL, between 1998 and 2017. Pathologically confirmed MSA was diagnosed by a single neuropathologist (D.W.D.) as previously described.⁵ Immunohistochemistry for α -synuclein (NACP, 1:3,000 rabbit polyclonal) was performed on sections of the basal forebrain, striatum, midbrain, pons, medulla, and cerebellum to establish a neuropathologic diagnosis. MSA cases were pathologically subclassified as MSA with predominant striatonigral involvement (MSA-P), MSA with predominant olivopontocerebellar involvement (MSA-C), and MSA with equally severe involvement of striatonigral and

olivopontocerebellar systems (MSA-mixed).⁵ One case was not assigned pathologic subtype because only the pons was available for histology.

The clinically diagnosed patients (N = 81 probable MSA, N = 8 possible MSA) were seen at the Mayo Clinic in Rochester, MN (N = 35), or Jacksonville, FL (N = 54), where diagnosis was made according to the criteria of Gilman et al.⁶ Clinically diagnosed MSA cases were free of notable cognitive symptoms that might suggest the presence of an overlapping dementia or other neurodegenerative conditions. Pathologic assessment and clinical diagnoses were made without the knowledge of the TREM2 p.R47H genotype. Controls were also from the Mayo Clinic in Rochester, MN (N = 991), or Jacksonville, FL (N = 704). Controls were free of neurologic symptoms and without a family history of movement disorders. All participants were unrelated non-Hispanic Caucasians. The primary comparison of TREM2 p.R47H involved the patients with pathologically confirmed MSA and controls. In secondary analysis, we combined the patients with pathologically confirmed MSA and patients with clinically diagnosed MSA and compared this combined MSA series with the controls. Table 1 shows characteristics of patients with MSA and controls.

Standard protocol approvals, registrations, and patient consents

The Mayo Clinic IRB approved the use of human participants in this study.

Genetic analysis

Genomic DNA was extracted from brain tissue or peripheral blood monocytes using standard protocols. Genotyping of *TREM2* exon 2 variant rs75932628 (NM_018965.3: c.140G>A, p.R47H) was performed using a custom TaqMan Allelic Discrimination Assay on an ABI 7900HT Fast Real-Time PCR system (Applied Bio-systems, Foster City, CA) according to the manufacturer's instructions (sequences available upon request). Genotype calls were made using SDS 2.2.2 software. Positive or ambiguous results in the TaqMan assay were also confirmed and resolved via Sanger sequencing. There was no evidence of a departure from Hardy-Weinberg equilibrium in controls ($p = 0.93$).

Statistical analysis

We evaluated the association between TREM2 p.R47H and MSA using logistic regression models. Unadjusted models were first examined. Subsequently, although it is uncertain whether adjusting for any variable is reasonable, given the rare nature of this variant, we did also adjust our logistic regression models for age and sex in a sensitivity analysis. Odds ratios (ORs) and 95%

Table 1 Characteristics of patients with MSA and controls

Variable	Patients with pathologically confirmed MSA (N = 168)	Patients with clinically diagnosed MSA (N = 89)	Controls (N = 1,695)
Age at onset in patients with MSA or age at blood draw in controls (y)	58 (39, 88)	64 (46, 83)	76 (30, 88)
Age at death (y)	66 (47, 91)	NA	NA
Sex			
Male	96 (57.1%)	53 (59.6%)	786 (46.4%)
Female	72 (42.9%)	36 (40.4%)	909 (53.6%)
MSA subtype			
MSA-P	73 (43.5%)	62 (69.7%)	NA
MSA-C	27 (16.1%)	26 (29.2%)	NA
Mixed ^a	67 (39.9%)	1 (1.1%)	NA
Unclassified ^b	1 (0.6%)	0 (0.0%)	NA
Braak NFT stage			
0	29 (17.8%)	NA	NA
I	61 (37.4%)	NA	NA
II	49 (30.1%)	NA	NA
III	16 (9.8%)	NA	NA
IV	5 (3.1%)	NA	NA
V	2 (1.2%)	NA	NA
VI	1 (0.6%)	NA	NA
Thal amyloid phase			
0	88 (55.7%)	NA	NA
1	33 (20.9%)	NA	NA
2	18 (11.4%)	NA	NA
3	9 (5.7%)	NA	NA
4	8 (5.1%)	NA	NA
5	2 (1.3%)	NA	NA

Abbreviations: MSA = multiple system atrophy; MSA-C = MSA with predominant olivopontocerebellar involvement; MSA-P = MSA with predominant striatonigral involvement; NA = not applicable.

The sample median (minimum, maximum) is given for age. Information was unavailable regarding age at onset (N = 49), Braak NFT stage (N = 5), and Thal amyloid phase (N = 10) for the patients with pathologically confirmed MSA.

^a MSA with equally severe involvement of striatonigral and olivopontocerebellar systems was classified as MSA-mixed.⁵

^b One patient with MSA was considered to be "unclassified" because only the pons was available for histologic assessment; therefore, we could not diagnose this patient as MSA-C or MSA-mixed.

confidence intervals were estimated. $p \leq 0.05$ was considered statistically significant. All statistical analyses were performed using SAS (version 9.4; SAS Institute, Inc., Cary, NC).

Results

An evaluation of the association between TREM2 p.R47H and MSA is shown in table 2. The p.R47H variant was observed in 3 patients with pathologically confirmed MSA (1.79%). Of these 3 p.R47H carriers, neuropathologic examination was available for 2 and revealed MSA subtypes of MSA-P and MSA-C, Braak neurofibrillary tangle stages of I and III, and Thal amyloid phases of 0 and 0. For the 89 patients with clinically diagnosed MSA, there was 1 p.R47H carrier (1.12%, from Mayo Jacksonville), and this patient's MSA subtype was probable MSA-P. There were 7 carriers of p.R47H in the controls (0.41%, N = 4 Mayo Jacksonville, N = 3 Mayo Rochester), corresponding to a minor allele frequency of 0.21%, which is similar to the frequency reported in the ExAC database (Cambridge, MA [exac.broad-institute.org]) for this population (0.26%).⁷ For the primary comparison of patients with pathologically confirmed MSA and controls, risk of MSA was significantly higher for p.R47H carriers (OR = 4.39, $p = 0.033$). When combining the pathologically confirmed and clinically diagnosed MSA series, the risk of MSA (vs controls) was also significantly higher (OR = 3.81, $p = 0.034$). In a sensitivity analysis adjusting for age and sex, the findings regarding the patients with pathologically confirmed MSA (OR = 3.55, $p = 0.076$) and the combined MSA series (OR = 3.17, $p = 0.078$) remained relatively consistent though weakened slightly and were no longer quite significant.

Discussion

TREM2 p.R47H is a well-established risk factor for AD⁴; however, results have been mixed for FTD and PD,⁴ while studies involving amyotrophic lateral sclerosis and dementia with Lewy bodies mostly point to a lack of association.⁴ Of interest, the results of our current study suggest that the TREM2 p.R47H variant may be associated with an increased risk of MSA. With a notably higher frequency in our combined MSA series (1.6%) compared with controls (0.4%), we observed a nearly 4-fold increased risk of MSA in p.R47H carriers. It is important that this finding was strongest in our pathologically confirmed MSA series. The absence of notable AD pathology in pathologically confirmed MSA p.R47H carriers indicates that the observed association was not driven by such pathology. The association was attenuated slightly when adjusting for age and sex; however, those results should not be overinterpreted because it is uncertain whether attempting any adjustment is reasonable, given the low frequency of p.R47H. These preliminary findings suggest that TREM2 p.R47H may play a role in susceptibility to MSA and that further study is warranted.

To the best of our knowledge, only 1 association study between TREM2 p.R47H and MSA has been conducted to date. In a study of 407 Chinese patients with clinically diagnosed MSA and 869 controls, the p.R47H substitution was observed in 1 MSA patient and no controls ($p = 0.14$).⁸ This negative result is not surprising because TREM2 p.R47H is extremely rare in East Asians.

Table 2 Association between TREM2 p.R47H and MSA

Disease group	N	No. (%) with TREM2 p.R47H	Unadjusted analysis		Adjusting for age and sex	
			OR (95% CI)	p Value	OR (95% CI)	p Value
Controls	1,695	7 (0.41)	1.00 (reference)	NA	1.00 (reference)	NA
Patients with pathologically confirmed MSA	168	3 (1.79)	4.39 (1.12–17.12)	0.033	3.55 (0.88–14.38)	0.076
All patients with MSA (pathologically confirmed and clinically diagnosed)	257	4 (1.56)	3.81 (1.11–13.12)	0.034	3.17 (0.88–11.45)	0.078

Abbreviations: CI = confidence interval; MSA = multiple system atrophy; NA = not applicable; OR = odds ratio; TREM2 = triggering receptor expressed on myeloid cells.

ORs, 95% CIs, and *p* values result from logistic regression models.

For age, the age that was adjusted for was age at death for patients with pathologically confirmed MSA, age at MSA onset for patients with clinically diagnosed MSA, and age at blood draw for controls.

How the TREM2 p.R47H variant could increase risk of MSA is an important topic for further study. MSA is regarded as an oligodendrogliaopathy, but the mechanism of demyelination remains unclear. While demyelination depends primarily on oligodendrocytes, microglia also contribute to myelination and myelin homeostasis.⁹ In the CNS, TREM2 is predominantly expressed in microglia, and it has been shown to play critical roles in phagocytic clearance of apoptotic cells and disease-associated molecules and modulating microglial immune response.¹⁰ A recent study showed that TREM2 is required for myelin debris removal and remyelination after cuprizone-induced demyelination.¹¹ We hypothesize that TREM2 p.R47H, a loss-of-function mutation, leads to impaired clearance of myelin debris and aberrant microglial activation, which result in increased risk of MSA.

Our results provide evidence that the TREM2 p.R47H substitution may be a genetic susceptibility factor for MSA. Our study is limited by the relatively small number of patients with pathologically confirmed MSA, and therefore, validation will be crucial. Recently, the clinical diagnostic accuracy of MSA was reported to be 62%, further underscoring the importance of using neuropathologically confirmed MSA cases in research studies.¹² We hope that these initial promising results will form the basis for future examinations of TREM2 p.R47H in MSA.

Author contributions

K. Ogaki: drafting/revising the manuscript; study concept and design; and interpretation of data. M. G. Heckman and S. Koga: drafting/revising the manuscript and interpretation of data. Y.A. Martens: drafting/revising the manuscript. C. Labbé and O. Lorenzo-Betancor: revising the manuscript. R.L. Walton, A.I. Soto, and E.R. Vargas: revising the manuscript and interpretation of data. S. Fujioka: revising the manuscript. R.J. Uitti, J. Van Gerpen, and W.P. Cheshire: revising the manuscript and providing material for this study. S.G. Younkin: revising the manuscript. Z.K. Wszolek: revising the manuscript, providing material and funding for this study, and IRB committee approval. P.A. Low and W. Singer: revising

the manuscript and providing material for this study. G. Bu: revising the manuscript. D.W. Dickson: drafting/revising the manuscript, providing material and funding for this study, and IRB committee approval. O.A. Ross: drafting/revising the manuscript; study concept and design; and interpretation of data.

Acknowledgment

The authors thank all those who have contributed to our research, particularly the patients and families who donated Brain and DNA samples for this work.

Study funding

K. Ogaki is supported by a research grant from the NAITO Foundation, JSPS KAKENHI Grant Number JP17K14966 and a grant from Institute for Environmental and Gender-Specific Medicine, Juntendo University. R.J. Uitti, O.A. Ross, Z.K. Wszolek, and D.W. Dickson are partially supported by the NIH/NINDS P50 NS072187. O.A. Ross is supported by the NINDS R01# NS078086, U54 NS100693, DOD W81XWH-17-1-0249, and in part by the Michael J. Fox Foundation for Parkinson's Research. C. Labbé is the recipient of a F.R.S.Q. fellowship. Z.K. Wszolek receives research support from NIH/NIA (primary), and NIH/NINDS (secondary) 1U01AG045390-01A1, Mayo Clinic Center for Regenerative Medicine, Mayo Clinic Neuroscience Focused Research Team (Cecilia and Dan Carmichael Family Foundation, and the James C. and Sarah K. Kennedy Fund for Neurodegenerative Disease Research at Mayo Clinic in Florida), and is also partially supported by a gift from Carl Edward Bolch, Jr, and Susan Bass Bolch, The Sol Goldman Charitable Trust, and Donald G. and Jodi P. Heeringa. P.A. Low receives research support from the Cure MSA Foundation. Samples included in this study were clinical patients or brain donors to the brain bank at Mayo Clinic in Jacksonville, which is supported by the CurePSP Society for Progressive Supranuclear Palsy and Udall Center for Excellence in Parkinson Research (P50 NS072187), APDA Center for Advanced Research, an Alzheimer's disease Research Center grant (NIA P50 AG16574), and NINDS R01 NS078086.

Disclosure

K. Ogaki receives JSPS KAKENHI Grant Number JP17K14966 and a grant from Institute for Environmental & Gender-Specific Medicine, Juntendo University. M.G. Heckman serves as the editorial board member of *Parkinsonism & Related Disorders*. S. Koga and Y.A. Martens report no disclosures. C. Labbé receives a Fonds de recherche du Québec-Santé postdoctoral fellowship. O. Lorenzo-Betancor, R.L. Walton, A.I. Soto, and E. Vargas report no disclosures. S. Fujioka receives JSPS KAKENHI Grant Number JP15K19501 and is an editorial board member of *Parkinsonism & Related Disorders*. R.J. Uitti serves as an associate editor of *Neurology*. J. Van Gerpen report no disclosures. W.P. Cheshire serves in the editorial boards of *Parkinsonism & Related Disorders* and *Autonomic Neuroscience* and as an associate editor of *Clinical Autonomic Research*. S.G. Younkin reports no disclosures. Z.K. Wszolek receives research support from P50 NS072187, NIH/NIA (primary), and NIH/NINDS (secondary) 1U01AG045390-01A1, Mayo Clinic Center for Regenerative Medicine, Mayo Clinic Neuroscience Focused Research Team (Cecilia and Dan Carmichael Family Foundation, and the James C. and Sarah K. Kennedy Fund for Neurodegenerative Disease Research at Mayo Clinic in Florida), the gift from Carl Edward Bolch, Jr., and Susan Bass Bolch, The Sol Goldman Charitable Trust, and Donald G. and Jodi P. Heeringa; serves as coeditor-in-chief of *Parkinsonism & Related Disorders* and an associate editor of the *European Journal of Neurology*; is on the editorial boards of *Neurologia i Neurochirurgia Polska*, *European Journal of Clinical and Experimental Medicine*, *Clinical and Experimental Medical Letters*, and *Wiadomosci Lekarskie*; holds and has contractual rights for receipt of future royalty payments from patents for “A Novel Polynucleotide Involved in Heritable Parkinson’s Disease”; and receives royalties from publishing *Parkinsonism & Related Disorders* (Elsevier, 2016, 2017) and the *European Journal of Neurology* (Wiley Blackwell, 2016, 2017). P.A. Low receives research support from the NIH (P01 NS44233, U54 NS065736, R01 NS092625, and UL1 TR000135), FDA (R01 FD004789), Cure MSA Foundation, and Mayo Funds and

serves as a clinical editor of *Autonomic Neuroscience*. W. Singer reports no disclosures. G. Bu received support from P50AG016574, RF1AG051504, R01AG027924, R01AG035355, R01AG046205, and P01NS074969 and a grant from the Cure Alzheimer’s fund. D.W. Dickson receives support from P50-AG016574, P50-NS072187, P01-AG003949, and CurePSP: Foundation for PSP|CBD and Related Disorders. D.W. Dickson is an editorial board member of *Acta Neuropathologica*, *Annals of Neurology*, *Brain*, *Brain Pathology*, and *Neuropathology* and is editor-in-chief of the *American Journal of Neurodegenerative Disease* and *International Journal of Clinical and Experimental Pathology*. O.A. Ross received support from R01 NS078086, P50-NS072187, and U54 NS100693 and the Michael J. Fox Foundation. O.A. Ross is an editorial board member of the *American Journal of Neurodegenerative Disease*, *Neurologia i Neurochirurgia Polska*, and *Molecular Neurodegeneration*. Full disclosure form information provided by the authors is available with the full text of this article at Neurology.org/NG.

Received January 23, 2018. Accepted in final form May 23, 2018.

References

1. Ahmed Z, Asi YT, Sailer A, et al. The neuropathology, pathophysiology and genetics of multiple system atrophy. *Neuropathol Appl Neurobiol* 2012;38:4–24.
2. Scholz SW, Bras J. Genetics underlying atypical parkinsonism and related neurodegenerative disorders. *Int J Mol Sci* 2015;16:24629–24655.
3. Sailer A, Scholz SW, Nalls MA, et al. A genome-wide association study in multiple system atrophy. *Neurology* 2016;87:1591–1598.
4. Jay TR, von Saucken VE, Landreth GE. TREM2 in neurodegenerative diseases. *Mol Neurodegener* 2017;12:56.
5. Koga S, Parks A, Uitti RJ, et al. Profile of cognitive impairment and underlying pathology in multiple system atrophy. *Mov Disord* 2017;32:405–413.
6. Gilman S, Wenning GK, Low PA, et al. Second consensus statement on the diagnosis of multiple system atrophy. *Neurology* 2008;71:670–676.
7. Lek M, Karczewski KJ, Minikel EV, et al. Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 2016;536:285–291.
8. Chen Y, Chen X, Guo X, et al. Assessment of TREM2 rs75932628 association with Parkinson’s disease and multiple system atrophy in a Chinese population. *Neurol Sci* 2015;36:1903–1906.
9. Simons M, Lyons DA. Axonal selection and myelin sheath generation in the central nervous system. *Curr Opin Cel Biol* 2013;25:512–519.
10. Painter MM, Atagi Y, Liu CC, et al. TREM2 in CNS homeostasis and neurodegenerative disease. *Mol Neurodegener* 2015;10:43.
11. Poliani PL, Wang Y, Fontana E, et al. TREM2 sustains microglial expansion during aging and response to demyelination. *J Clin Invest* 2015;125:2161–2170.
12. Koga S, Aoki N, Uitti RJ, et al. When DLB, PD, and PSP masquerade as MSA: an autopsy study of 134 patients. *Neurology* 2015;85:404–412.

Neurology® Genetics

Association study between multiple system atrophy and TREM2 p.R47H

Kotaro Ogaki, Michael G. Heckman, Shunsuke Koga, et al.

Neurol Genet 2018;4;

DOI 10.1212/NXG.0000000000000257

This information is current as of August 1, 2018

Neurol Genet is an official journal of the American Academy of Neurology. Published since April 2015, it is an open-access, online-only, continuous publication journal. Copyright Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology.. All rights reserved. Online ISSN: 2376-7839.

Updated Information & Services	including high resolution figures, can be found at: http://ng.neurology.org/content/4/4/e257.full.html
References	This article cites 12 articles, 0 of which you can access for free at: http://ng.neurology.org/content/4/4/e257.full.html##ref-list-1
Citations	This article has been cited by 1 HighWire-hosted articles: http://ng.neurology.org/content/4/4/e257.full.html##otherarticles
Subspecialty Collections	This article, along with others on similar topics, appears in the following collection(s): All Clinical Neurology http://ng.neurology.org/cgi/collection/all_clinical_neurology All Genetics http://ng.neurology.org/cgi/collection/all_genetics All Movement Disorders http://ng.neurology.org/cgi/collection/all_movement_disorders Multiple system atrophy http://ng.neurology.org/cgi/collection/multiple_system_atrophy
Permissions & Licensing	Information about reproducing this article in parts (figures,tables) or in its entirety can be found online at: http://ng.neurology.org/misc/about.xhtml#permissions
Reprints	Information about ordering reprints can be found online: http://ng.neurology.org/misc/addir.xhtml#reprintsus

Neurol Genet is an official journal of the American Academy of Neurology. Published since April 2015, it is an open-access, online-only, continuous publication journal. Copyright Copyright © 2018 The Author(s). Published by Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology.. All rights reserved. Online ISSN: 2376-7839.